



# BlueMarine<sup>3</sup>.Com

Developing shrimp  
larval rearing  
techniques for  
application in Europe

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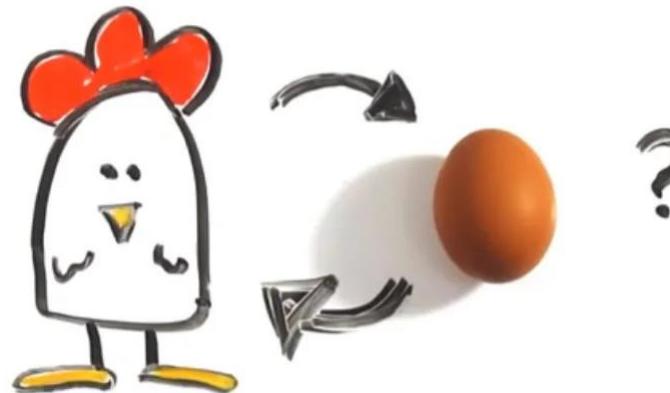


AGENTSCHAP  
INNOVEREN &  
ONDERNEMEN



# Context

- Growing interest in aquaculture/restoration
  - Roadmap/desk top studies: Aquavalue, SeaCconomy, ...
  - Nearshore / offshore projects: Value@sea, Edulis, Coastbusters, Symapa, United, Wier en Wind,...
  - Onland: shrimp, seaweed, ...
- Common problem: suitable starting material?
  - No marine hatchery in Belgium
  - Import often problematic:
    - Availability
    - Mortality during transport
    - Bio-security (SPF)
  - Lots of expertise, but lack of concerted approach



# General objectives

- Expand knowledge on hatchery techniques for 3 species groups
  - Adapt to local conditions (e.g. availability of water, energy, labour cost)
  - Specific local needs (species, genetic, disease status,...)
  - Increase ecological and economical sustainability
  - Taking advantage of specific activities of consortium partners
- No simple duplication, but also innovation
- Set up a (multispecies) hatchery pilot as an incubator / accelerator



# Specific goals

## WP2: Seaweed



## WP3: Mollusks



## WP4: Shrimp



## WP1: Synergies and integration?

- Hatchery-facilities
- Collection of local strains
- Life cycle control
- Genetic map / strain selection Ulva
- Provide starting material
- Bio-degradable substrate + seeding technique

- Hatchery- and nursery- protocol for local conditions
- RAS /automation
- Functional algae-based feeds
- Disease control
- Spuikom as nursery

- Species selection
- Indigenous species
- Hatcheryprotocol for local conditions
- Functional algae-based feeds
- RAS / automation
- New disease testing tools

## WP5: Ecological and economical gain

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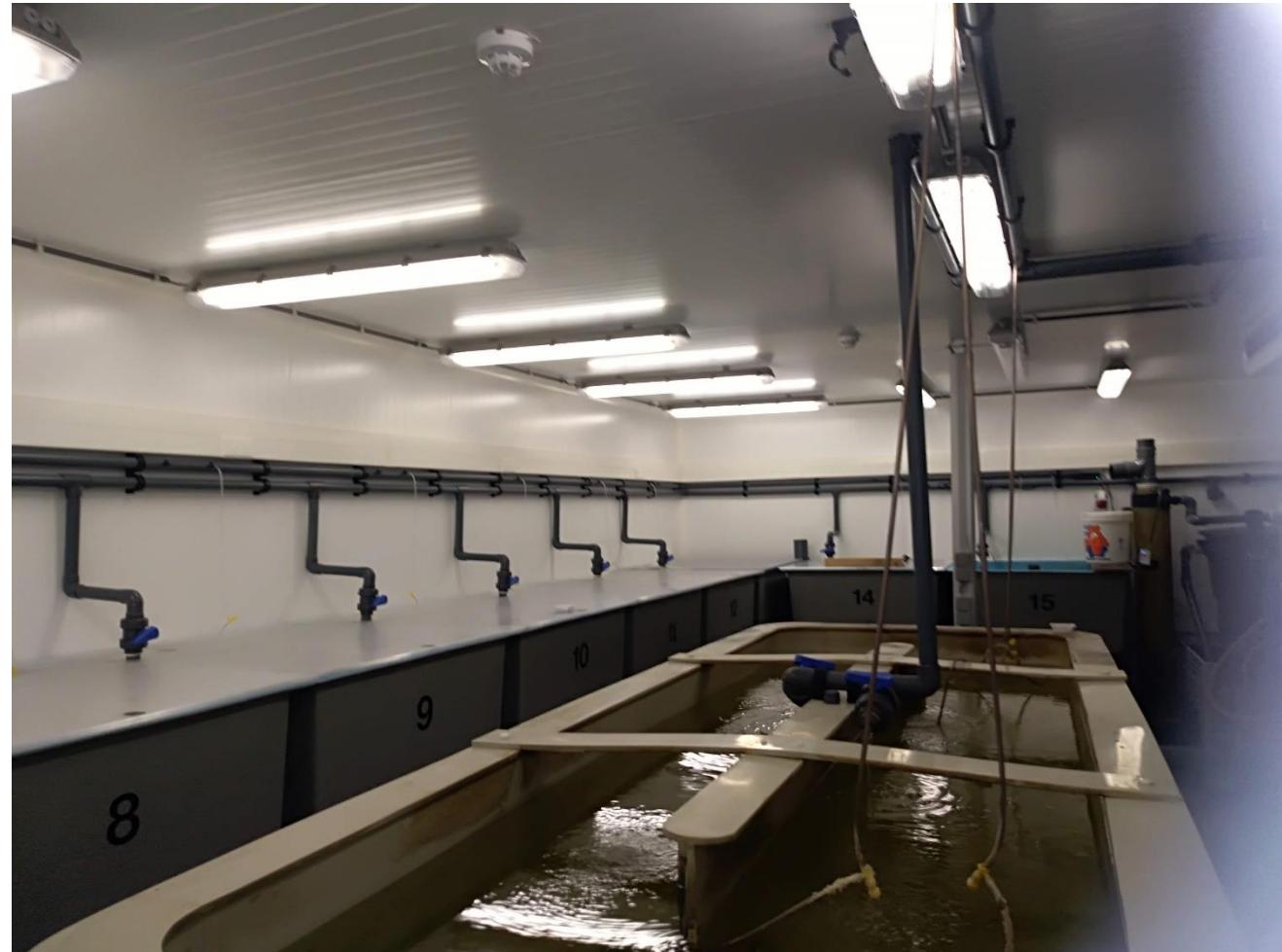
Blueprint for a viable hatchery of 3 marine commercially attractive species groups

- Flemish innovation and Entrepreneurship (Vlaio) - Blue Cluster - ICON-type project
  - Combination basic research / industrial research
  - Start: October 1, 2019; 39 months



# Availability of shrimp (*vannamei*) postlarvae

- Research at UGent (+others)
    - Thailand → Brazil → Thailand → US
  - Imaqua BVBA
  - Shrimp farming initiatives in Europe
- Not reliable, quality?
- Local production of PIs?



# First project results - shrimp

- ✓ Hatchery protocol *L. vannamei*
- ✓ Functional algae-based feeds



## Larval rearing system

### ➤ RAS set-up

- 12 tanks of 100 L
- Biofilter + protein skimmer
- Upflow - Flow rate adjustable
- Controlled room
  - 29 °C
  - 12L/12D

### ➤ Nauplii from IMAQUA BVBA

# Broodstock holding system and conditions

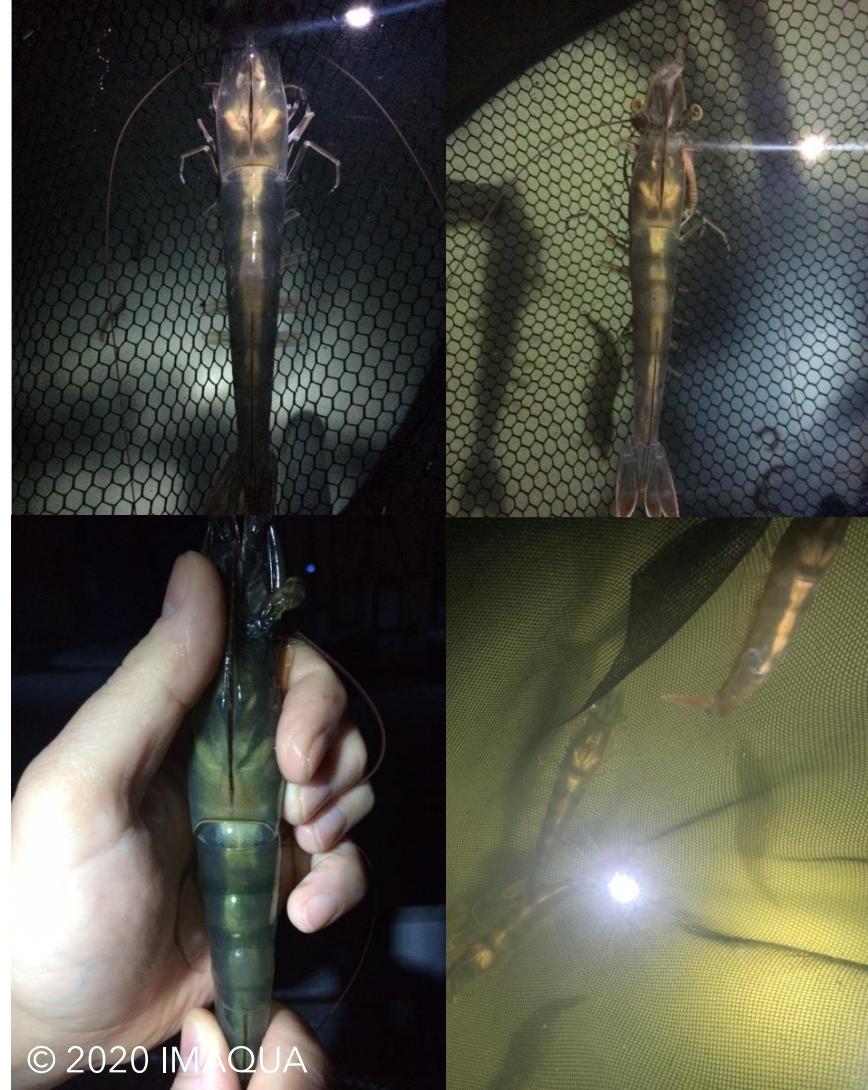
- 6 cylindrical tanks of 1.2 m<sup>3</sup>
- Biofilter, drumfilter, skimmer and UV
- Automatic feeding
- Inverted photoperiod
- Temp. 27°C, Salinity 32 ppt
- Demineralised water + commercial sea salt-mix
- Diet: IMAQUA broodstock dry feed (IM42), polychaetes and squid



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# Broodstock

- Origin: USA
- First broodstock raised and matured from postlarvae in a nursery system at IMAQUA
- Currently preparing F2 broodstock generation



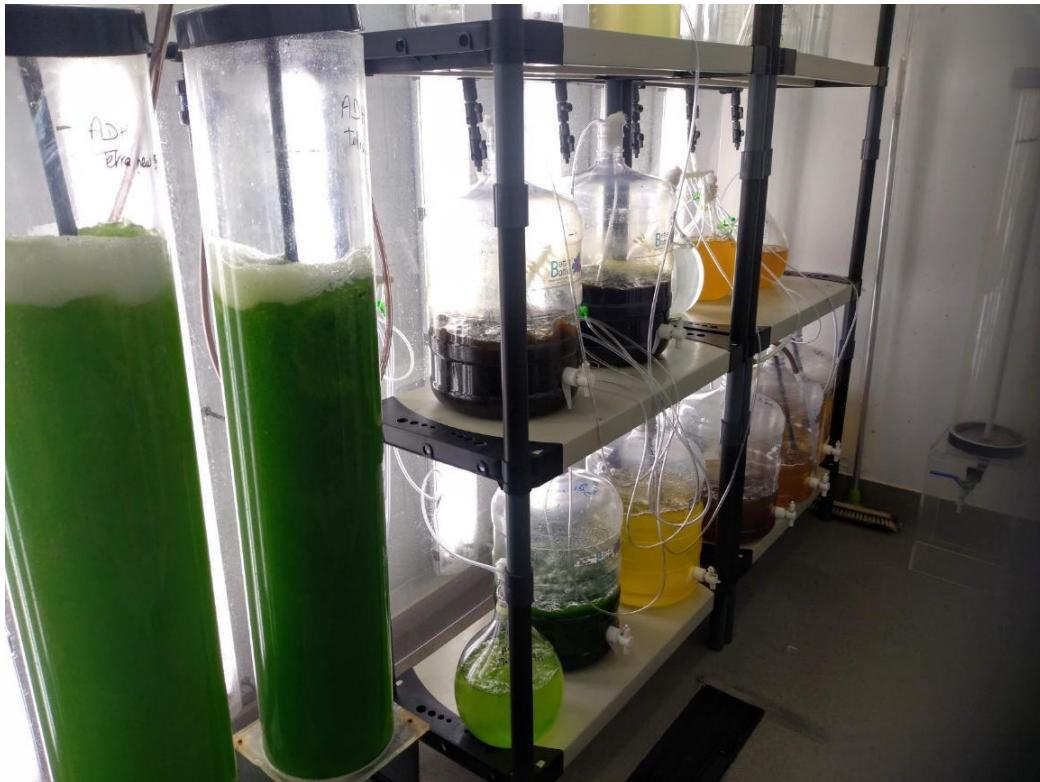
# Artificial insemination of females

- Allows to use smaller tanks and increase density of animals
- Most cost-effective option
- Best for European conditions
- Chosen as standard procedure at IMAQUA



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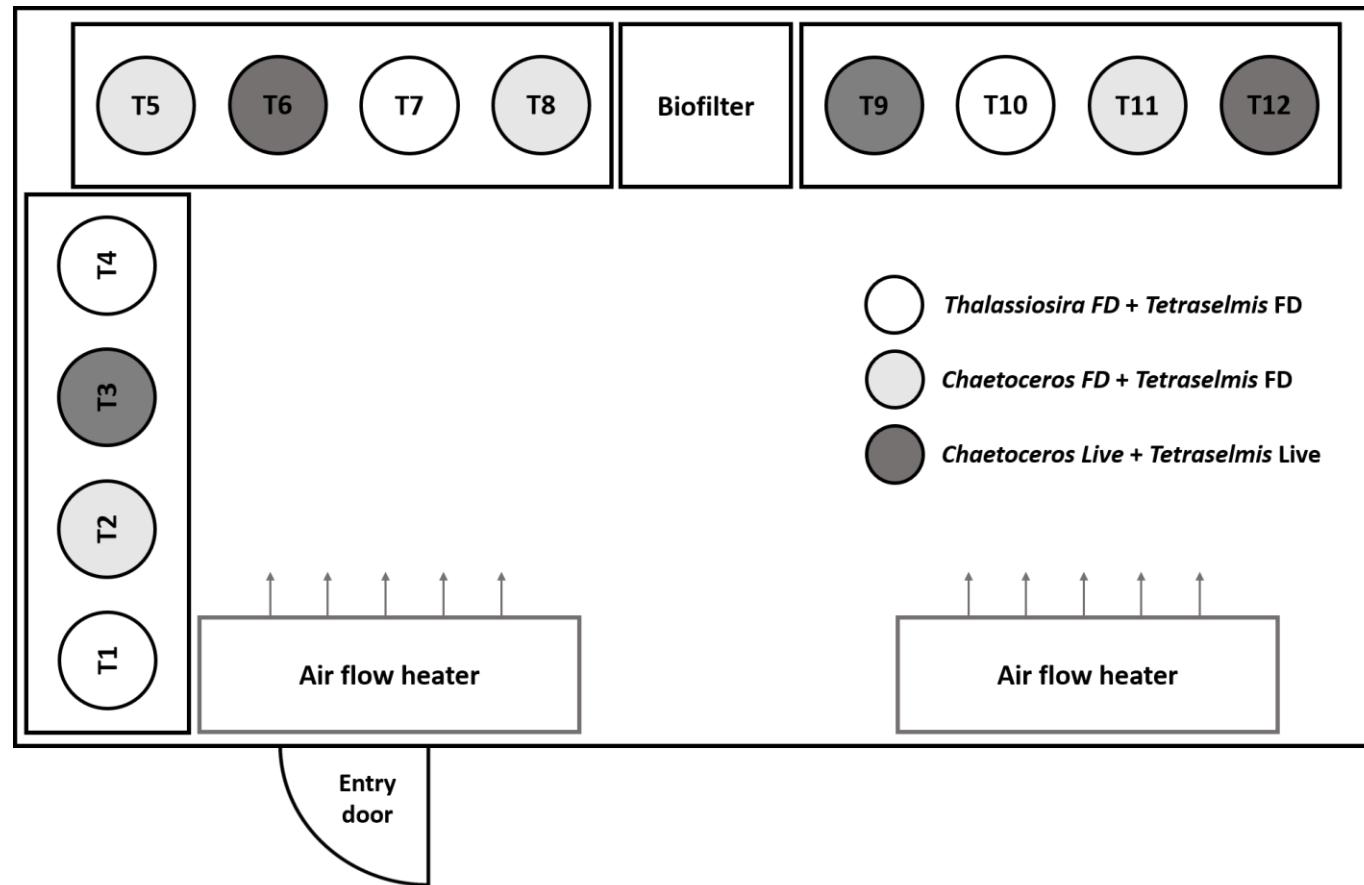
# Microalgae



- “In house” batch algae
- High-density photobioreactor algae (Proviron)



# Larval rearing protocol



- **Standard feeding schedule**
  - Algae  $\sim 10^5$  cells/ml
  - Artemia from Z3 onwards
- **Water regime**
  - Natural seawater
  - No exchange: N and Z stages, RAS from M
- **Parameters**
  - Residual algae → flow cytometry
  - Water quality parameters
  - Development and performance (activity, digestion and fouling)
  - Survival to PL stage
  - (Bacterial load)

# Larval rearing experiments

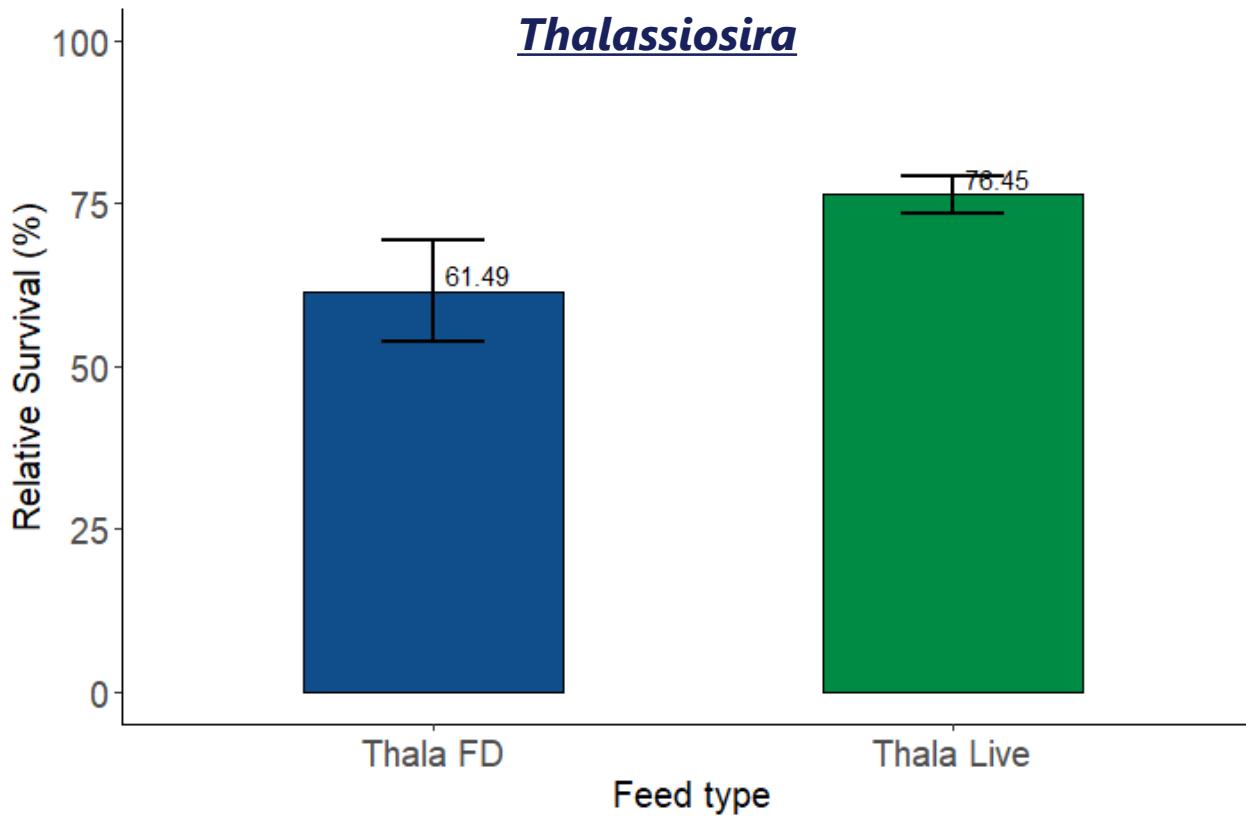
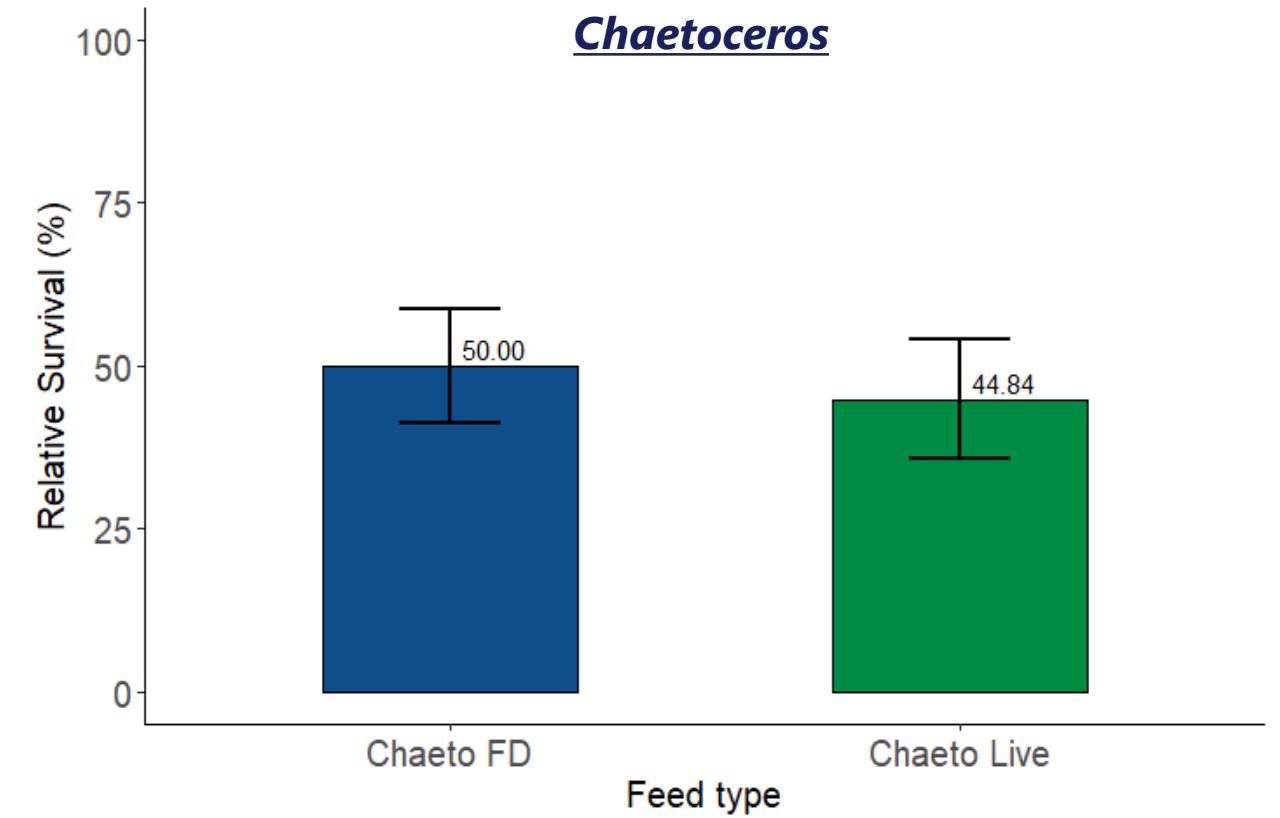
## ➤ Several factors tested

- Different mono- and mixed-species diets → **suitable algae species?**
- Batch- versus PBR algae → **effect culture technology algae?**
- Freeze-dried versus live algae → **effect processing algae?**
- Addition of probiotics grown on exudates of freeze-dried algae → **effect microflora?**

## ➤ Set-up/procedure systematically improved

- Fine-tuning aeration, turbulence, temperature and salinity control
- Biofilter efficiency
- ...

# Freeze-dried vs. live algae: Survival



# Freeze-dried vs. live algae: Larval development

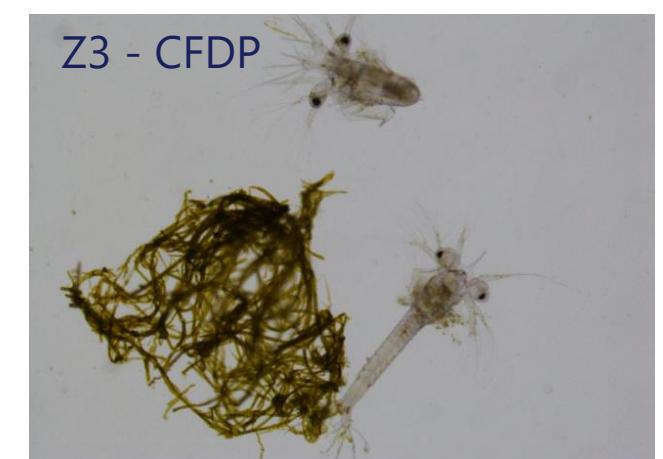
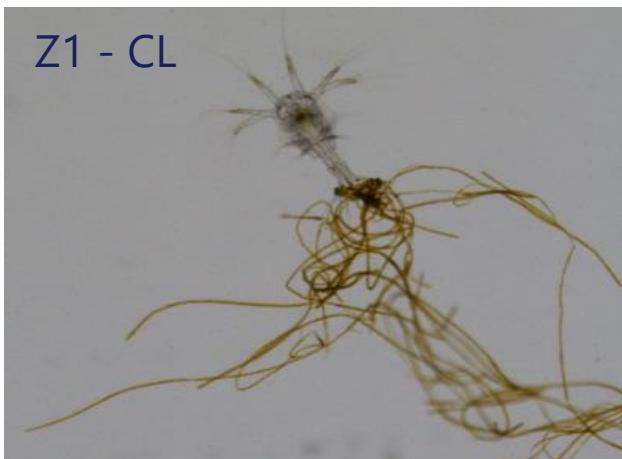
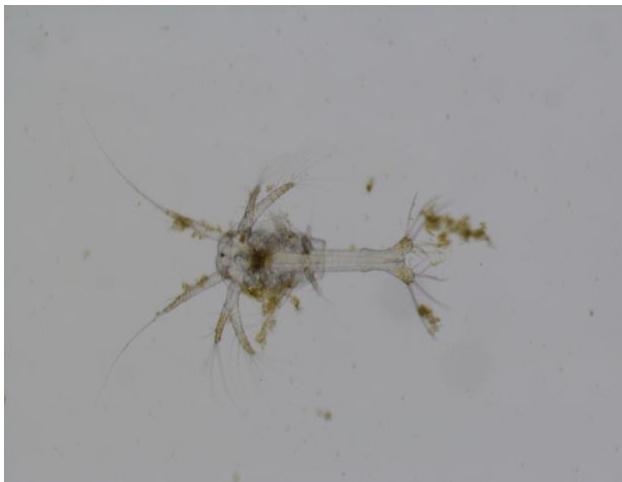
## Delay in development (1-1,5d) with freeze-dried algae

Day	CL	CLP	CFD	CFDP
0	N2-N3	N2-N3	N2-N3	N2-N3
1	N4-N5	N4-N5	N4-N5	N4-N5
2	Z1	Z1	Z1*	Z1*
3	Z2	Z2	Z2*	Z2*
4	Z2	Z2	Z2*	Z2*
5	<b>Z3</b>	<b>Z3</b>	<b>Z2*-Z3*</b>	<b>Z2*-Z3*</b>
6	M1	M1	Z3*-M1	Z3*-M1
7	M1-M2	M1-M2	M1-M2	M1
8	M2-M3	M2-M3	M2	M1-M2
9	M3- <b>PL1</b>	M3- <b>PL1</b>	M2-M3	M2-M3
10	PL1+	M3-PL1+	M3- <b>PL1</b>	M3- <b>PL1</b>
11	PL1+	PL1+	PL1+	PL1+

Day	TL	TFD
0	N2-N3	N2-N3
1	N4-N5	N4-N5
2	Z1	Z1*
3	Z2	Z2*
4	Z2	Z2*
5	<b>Z3</b>	<b>Z2*-Z3*</b>
6	Z3-M1	Z3*
7	M1-M2	Z3*-M1
8	M2-M3	M1-M2
9	M3- <b>PL1</b>	M2-M3
10	M3-PL1	M3- <b>PL1</b>
11	PL1+	M3-PL1

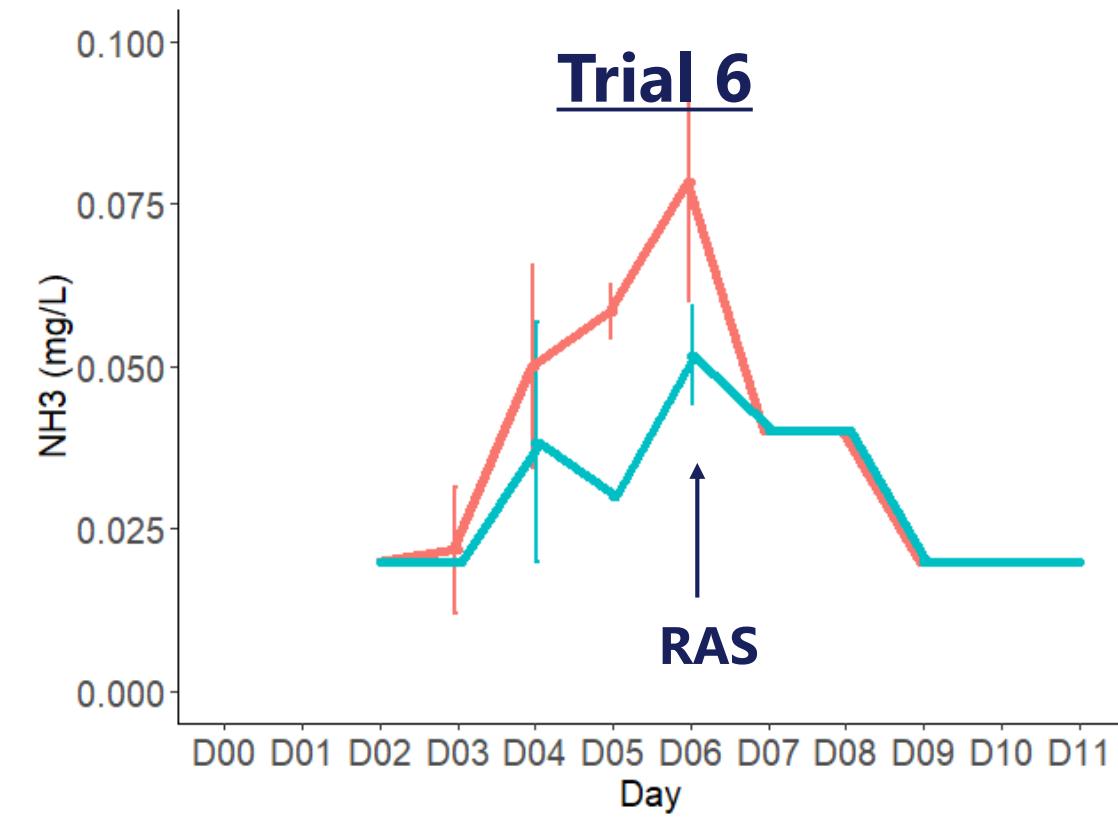
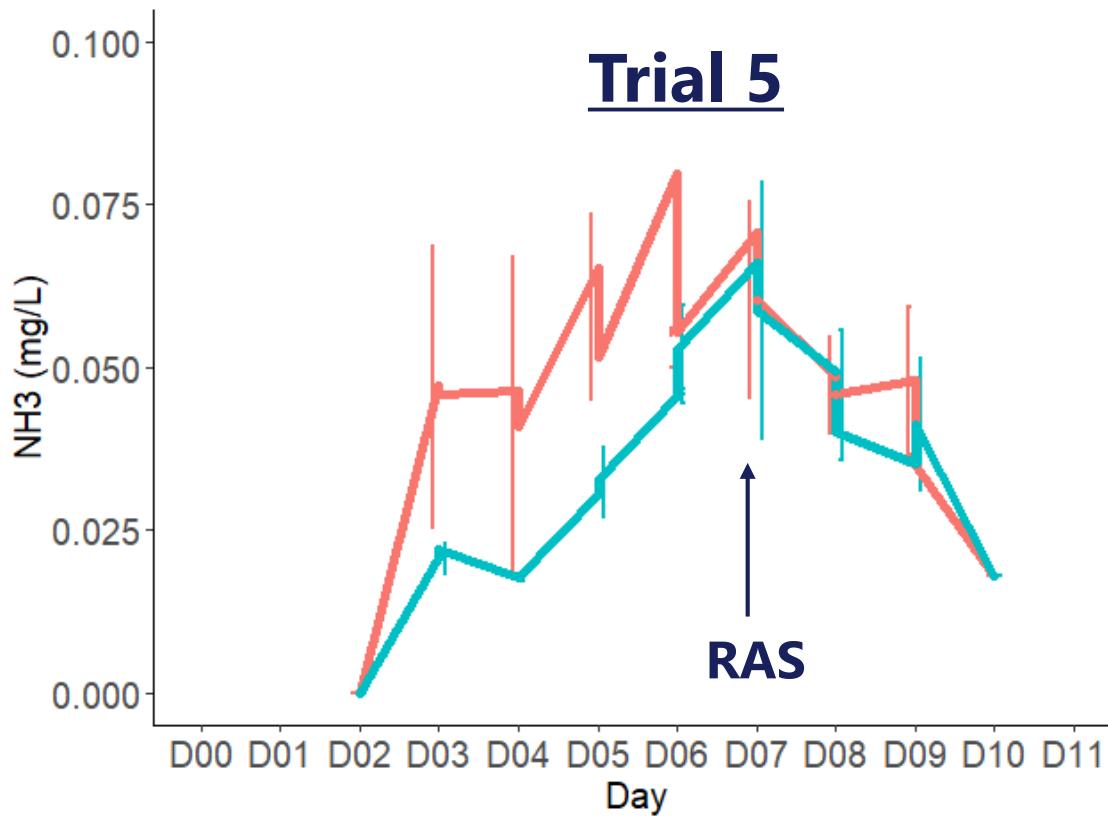
# Freeze-dried vs. live algae: Fouling and digestion

- Fouling: Freeze-dried algae
- "Lange faeces": both



# Freeze-dried vs. live algae: Water quality

- Temperature, salinity, DO → very stable
- NH<sub>3</sub> concentrations



# Conclusions hatchery protocol *L. vannamei*

## ➤ Good baseline

- Reliable and reproducible with live and freeze-dried algae
- Very low water consumption ( $\pm$  20L per day / 100,000 postlarvae)

## ➤ Next steps

- RAS from early stages (zoea): solution for water quality/fouling (with FD algae)?
  - Adjust feeding levels to compensate for losses in RAS
- Automatic algae feeding system: solution for overfeeding?
- Test alternative sea water sources (artificial, diluted brine): more widely applicable
- In parallel: probiotics grown on algae-exudates (C:N ratio)

## Thank You!

